

Stochastic models of protein production with cell division and gene replication

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Presentation

Biological context

Classical models for protein production

Model with cell division and gene replication

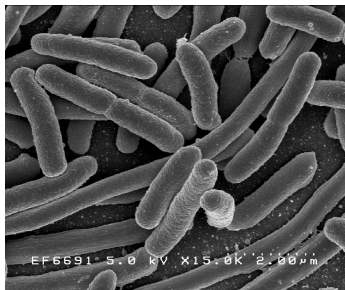
Results and further work

Part 1

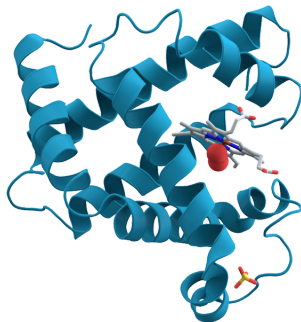
Biological context

Cells and proteins

- ▶ Cells: unit of life.
- ▶ Its goal: grow and divide.



- ▶ Functional molecules:
proteins
 - ▶ enzymes, wall, energy, etc.
- ▶ Produced from the genes



Protein production: A central mechanism

Proteins represents:

- ▶ 50% of the dry mass
- ▶ ~ 3 million molecules
- ▶ ~ 2000 different types
- ▶ from few dozens up to 10^5 proteins per type

It needs to be duplicated in one cell cycle (approx. 30 *min*)

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67% of the resources for protein production

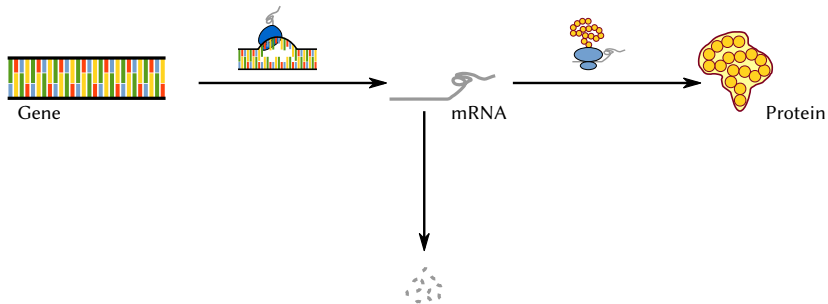
Classic protein production mechanism

Two main steps in protein production:

1. Transcription: to produce mRNA
2. Translation: to produce proteins

Transcription

Translation



Highly variable process

The protein production subject to high variability:

- ▶ Thermal noise (random collision between molecules)
- ▶ Cell events (division, gene replication)
- ▶ Fluctuations in of common resources

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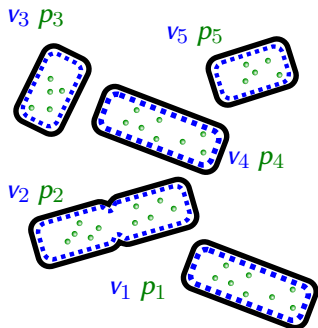
Problem: the main mechanism of the cell, impacted by a large variability.

**”How the cell deals with this variability?”
A main topic for experimental research.**

Taniguchi et al. (2010) experimental measures

Population of cells

- ▶ Measure volume v_i
- ▶ Measure of prot. number p_i



Interest in concentrations

Empirical mean:

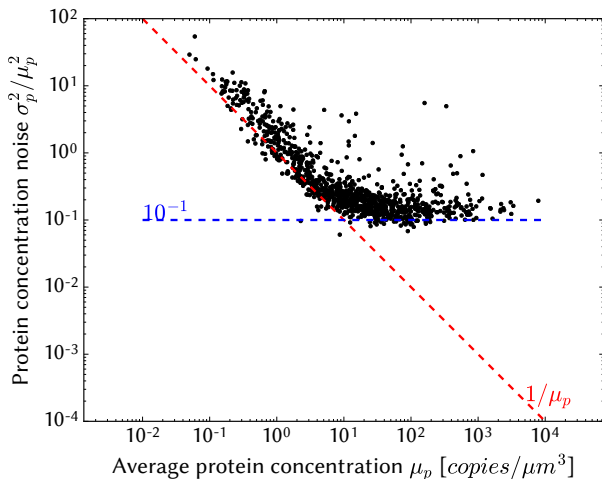
$$\mu_p = \frac{1}{N} \sum_{i=1}^N \frac{p_i}{v_i}$$

Empirical variance:

$$\sigma_p^2 = \frac{1}{N} \sum_{i=0}^N \left(\frac{p_i}{v_i} \right)^2 - \mu_p^2$$

Taniguchi et al. (2010) experimental measures

Two regimes in the protein variability:



Goal: modelling the protein production

- ▶ Models to describe the stochastic protein variability.
- ▶ Confront the models to real experiments (two regimes)

Part 2

Classical models for protein production

Markovian description

Framework for protein production modeling:

- ▶ Rigney and Schieve (1977)
- ▶ Berg (1978)
- ▶ Paulsson (2005)

Three types of events:

- ▶ Encounter between molecules
- ▶ mRNA and protein creation
- ▶ Lifetime of molecules

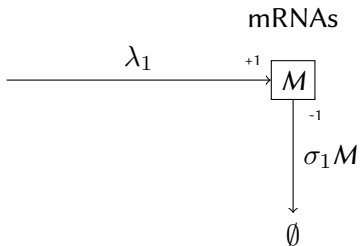
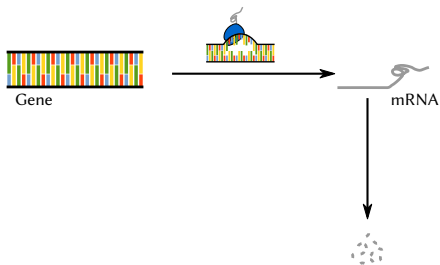
Assumption: Exponential times

Each event occurs at exponentially distributed time.

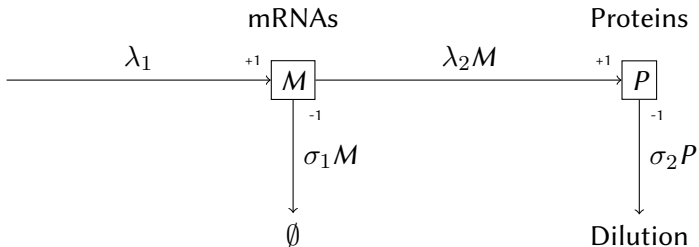
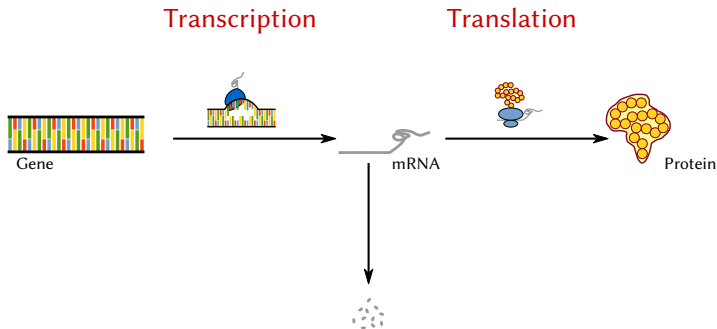
The classical model

The classical model

Transcription



The classical model



Limitations of classical models

Classical model, at equilibrium mean $\mathbb{E}[P]$ and the variance $\text{Var}[P]$ are known Paulsson (2005).

But this model has some limitations:

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We need to have a model with the notion of cell cycle.

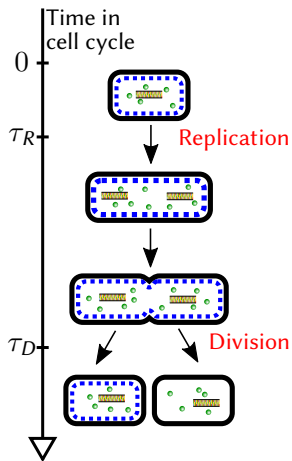
Part 3

Model with cell division and gene replication

Features of the model

A model with cell cycle:

- ▶ Considering a growing cell
- ▶ Gene replication at τ_R
- ▶ Division at τ_D

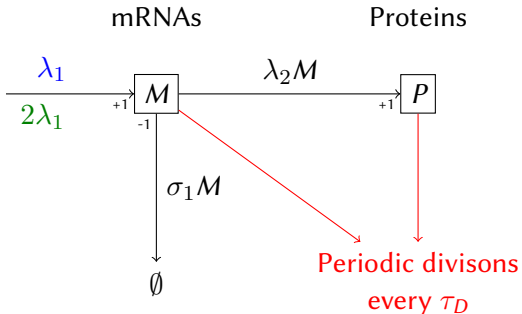


Times τ_R and τ_D are considered as deterministic.

Presentation of the model

Before replication:

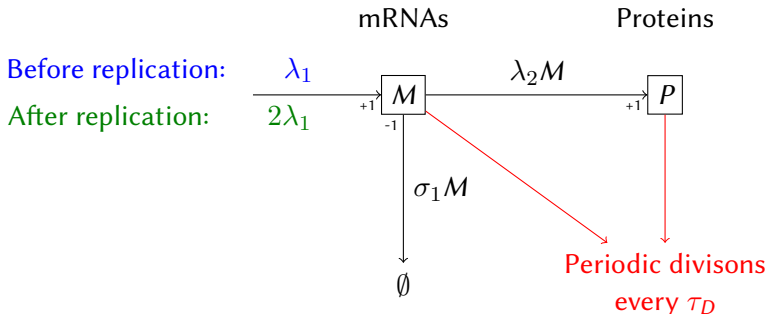
After replication:



Volume growth:

$$V(s) = V(0)2^{s/\tau_D}$$

Presentation of the model



Volume growth:

$$V(s) = V(0)2^{s/\tau_D}$$

Concentrations can be considered:

$$P_s/V(s) \quad \text{and} \quad M_s/V(s)$$

Explicit solution for the number of mRNAs

For any time s of the cell cycle the distribution of M_s is known.

Theorem

At equilibrium, at a time s in the cell cycle, the mRNA number M_s follows a Poisson distribution of parameter

$$x_s = \frac{\lambda_1}{\sigma_1} \left[1 - \frac{e^{-(s+\tau_D-\tau_R)\sigma_1}}{2 - e^{-\tau_D\sigma_1}} + \mathbb{1}_{s \geq \tau_R} \left(1 - e^{-(s-\tau_R)\sigma_1} \right) \right].$$

We need to use Marked Poisson Point Process for the proof.

Explicit solution for the mean and the variance

With more calculus, the first two moments of P_S are known.

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Theorem

At equilibrium, at any time s of the cell cycle, the mean and the variance of the protein number P_s are

$$\begin{aligned}\mathbb{E}[P_s] &= \lambda_2 (f_1(\tau_R) + f_2(\tau_D) + f_1(\tau_R \wedge s) + \mathbb{1}_{s \geq \tau_R} f_2(s)) \\ \text{Var}[P_s] &= \text{Var}[P_0] + 2\lambda_2 \frac{1 - e^{-\sigma_1 s \wedge \tau_R}}{\sigma_1} \text{Cov}[P_0, M_0] + g_1(s \wedge \tau_R) \\ &\quad + \mathbb{1}_{s \geq \tau_R} \left(2\lambda_2 \frac{1 - e^{-\sigma_1 (s - \tau_R)}}{\sigma_1} \text{Cov}[P_{\tau_R}, M_{\tau_R}] + g_2(s) \right)\end{aligned}$$

with $f_1, f_2, g_1, g_2, \text{Var}[P_0], \text{Cov}[P_0, M_0]$ and $\text{Cov}[P_{\tau_R}, M_{\tau_R}]$ explicitly depending on $\lambda_1, \sigma_1, \lambda_2, \tau_R$ and τ_D .

Part 4

Results and further work

Parameters

We use the empirical mean and variance of proteins in Taniguchi et al. (2010) to fit the parameters.

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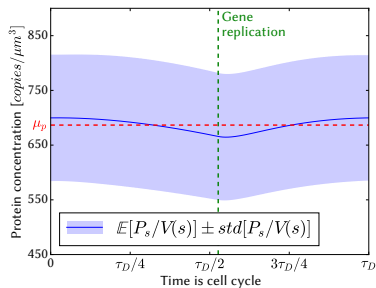
For each type of protein:

$$\mu_p = \frac{1}{\tau_D} \int_0^{\tau_D} \frac{\mathbb{E}[P_s]}{V(s)} ds.$$

Protein profile

The previous theorem can predict the protein variability:

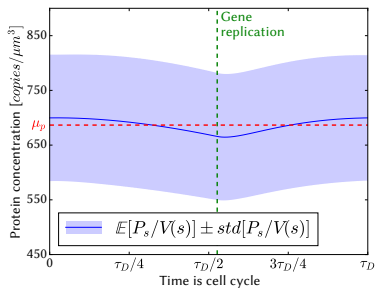
Simulations



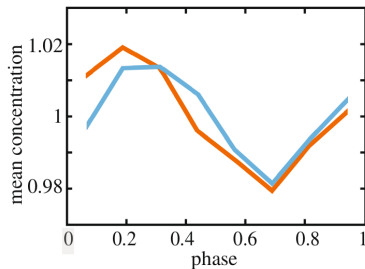
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Experiments

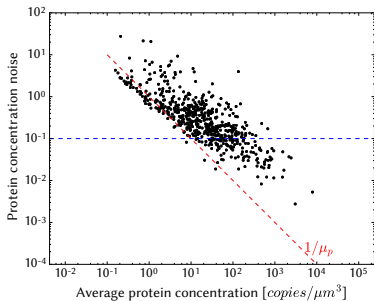


adapted from fig 4.b of Walker et al. (2016)

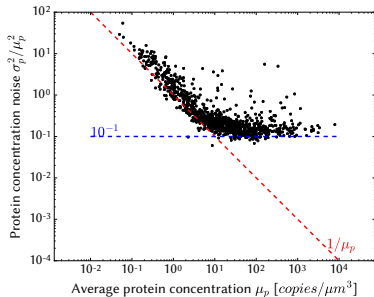
Protein noise

Direct comparison with Taniguchi et al. (2010)

Simulations



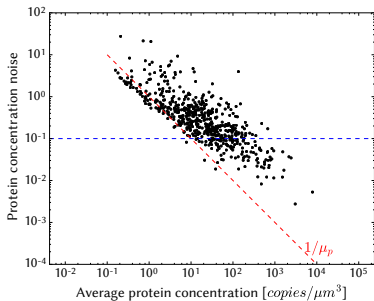
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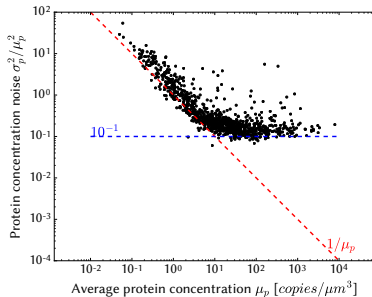
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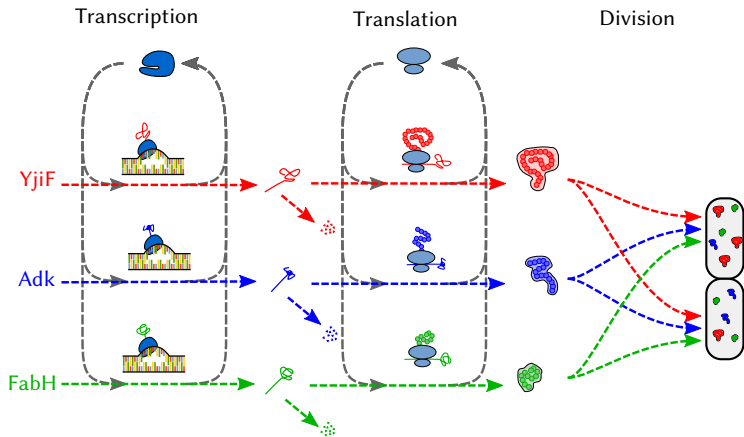
Experiments



A more complex model is needed

Multi-protein model

Model with a sharing of common resources: RNA-polymerases and ribosomes:



Conclusions

In this work:

- ▶ A model with division and replication
- ▶ Analytical results for protein mean and variances
- ▶ On average, coherent with experiments

But it does not reproduce all of the protein variability.

Thank you for you attention

PhD work supervised by

▶ Vincent Fromion



▶ Philippe Robert



For each gene, Taniguchi et al. (2010) gives:

- ▶ empirical mean of mRNA concentration: μ_m
 - ▶ empirical mean of protein concentration: μ_p
 - ▶ mRNA lifetime σ_1
 - ▶ gene position (from which τ_R can be deduced)

Main idea of the proof

Question: How many mRNAs X_s

- ▶ created since the birth of the cell
- ▶ still present at time s (with time s before replication)

Use of a Marked Poisson Point Process of intensity

$$\nu(dx, dy) = \lambda_1 dx \otimes \sigma_1 e^{-\sigma_1 y} dy.$$

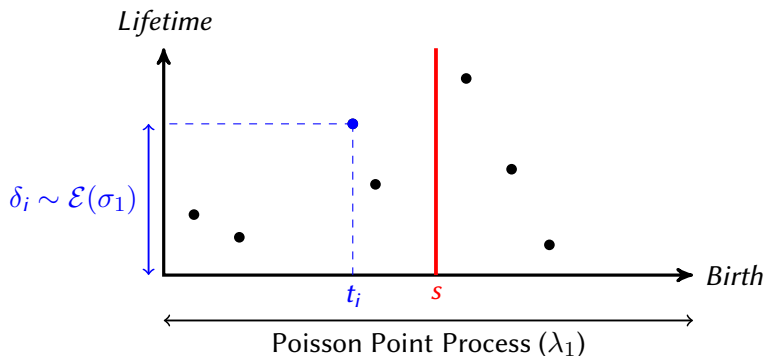
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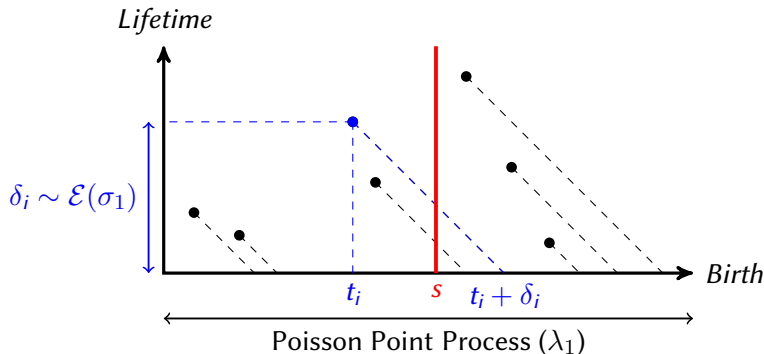
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